


new matter is introduced by virtue of these amendments, and the amendments are fully supported by the Specification of the subject application and the claims as originally filed. Accordingly, applicants kindly request that they be entered into the instant application.

**STATEMENT ACCOMPANYING SEQUENCE LISTING**

The undersigned hereby states that the Sequence Listing submitted concurrently herewith does not include matter which goes beyond the content of the application as filed and that the information recorded on the diskette submitted concurrently herewith is identical to the written Sequence Listing.

	Respectfully submitted,
Date      January 2, 2003	
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--In order to get the flanking sequences of the NS RNA fused to the coding sequence of the CAT gene, the following strategy was used. Two suitable internal restriction sites were selected, close to the start and stop codon of the CAT gene, that would allow the replacement of the sequences flanking the CAT gene in the pCM7 plasmid with the 3'- and 5'-NS RNA sequences. At the 5' end, a SfaNI site was chosen, (which generates a cut 57 nt from the ATG) and at the 3'- end a ScaI site which generates a cut 28 nt from the end of the gene (stop codon included). Next, four synthetic oligonucleotides were made using an Applied Biosystems DNA synthesizer, to generate two double-stranded DNA fragments with correct overhangs for cloning. Around the start codon these oligonucleotides formed a piece of DNA containing a XbaI overhang followed by a HgaI site and a PstI site, the 3'-(viral-sense) NS sequence immediately followed by the CAT sequence from start codon up to the SfaNI overhang (underscored). In addition a silent mutation was incorporated to generate an AccI site closer to the start codon to permit future modifications.

Around the stop codon the two other oligonucleotides generated a piece of DNA as follows: a blunt-ended ScaI site, the CAT sequence from this site up to and including the stop codon (underlined) followed by a BglII site and a Xba I overhang.

Sca I Bgl II  
5'-actgcgatgagtggcagggcgggcgtaatagat- 3' (SEQ ID NO: 22) oligo3

3'-tgacgctactcacggtcccgccccgcattatctagatc- 5' (SEO ID NO: 25)

oligo4

XbaI

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**Exhibit B**  
**Claims As Pending Upon Entry Of The Amendment**

35. A method for producing a chimeric negative strand RNA virus, comprising culturing a host cell transfected with plasmid cDNAs containing a heterologous nucleotide sequence operatively linked to a binding site specific for an RNA-directed RNA polymerase of a negative strand RNA virus, wherein the host cell expresses a polymerase proteins, and recovering a chimeric virus from culture.

36. The method of Claim 35 wherein the host cell constitutively expresses the polymerase proteins.

37. A chimeric virus recovered from the method of Claim 35.

38. A method for producing a chimeric negative strand RNA virus, comprising culturing a host cell transfected with plasmid DNAs containing a heterologous nucleotide sequence operatively linked to a binding site specific for an RNA-directed RNA polymerase of a negative strand RNA virus, and with plasmid DNAs containing nucleotide sequences which encode an RNA polymerase proteins, and recovering a chimeric virus from culture.

39. The method of Claim 38 wherein the chimeric virus is influenza virus.

40. The method of Claim 39 wherein the heterologous RNA segment is derived from another strain of influenza virus.

41. A chimeric virus recovered from the method of Claim 39.

42. A method for producing a chimeric negative strand RNA virus comprising culturing a host cell transfected with plasmid cDNAs containing the nucleotide sequences encoding eight genomic segments from different strains of influenza virus, each of the segments comprising the reverse complement of an mRNA coding sequence for an RNA-directed RNA polymerase of a negative strand virus, wherein the host cell expresses an RNA polymerase protein, and recovering a chimeric virus from culture.

43. The method of Claim 42 wherein the host cell constitutively expresses the polymerase proteins.

44. A chimeric virus recovered from the method of Claim 42.

45. A method of producing a chimeric negative strand RNA virus, comprising culturing a host cell transfected with plasmid cDNAs containing a heterologous nucleotide sequence comprising a sequence mutated from a wildtype sequence of the negative strand RNA virus, operatively linked to a binding site specific for an RNA-directed RNA polymerase of a negative strand RNA virus, wherein the host cell expresses a polymerase protein and recovering a chimeric virus from culture.

46. The method of claim 45 wherein the sequence mutated is a site specific mutation.

47. The method of claim 45 wherein the virus is influenza.

-- (New) 48. A method for producing a negative strand RNA virus, comprising culturing a host cell transfected with plasmid cDNAs containing a nucleotide sequence operatively linked to a binding site specific for an RNA-directed RNA polymerase of a negative strand RNA virus, wherein the host cell expresses a polymerase proteins, and recovering a virus from culture.

(New) 49. The method of Claim 48 wherein the host cell constitutively expresses the polymerase proteins.

(New) 50. A method for producing a negative strand RNA virus, comprising culturing a host cell transfected with plasmid DNAs containing a nucleotide sequence operatively linked to a binding site specific for an RNA-directed RNA polymerase of a negative strand RNA virus, and with plasmid DNAs containing nucleotide sequences which encode an RNA polymerase proteins, and recovering a virus from culture.

(New) 51. The method of Claim 48 or 50 wherein the virus is influenza virus.

(New) 52. The method of Claim 48 or 50 wherein the plasmid DNA contains a heterologous nucleotide sequence.

(New) 53. A virus recovered from the method of Claim 52. --